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SYNTHESIS AND ANTITUMOUR PROPERTIES OF 2-THIO-5-CHLORO-NUCLEOSIDES

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Abstract: Four methods are described for the synthesis of 2-thio-5-chlorouracil (1). β - and α -5-Chloro-2-thio-2'-deoxyuridines (12 and 13) were obtained by Lewis acid catalysed condensation of TMS derivative of 1 with 2-deoxy-3,5-di-O-p-toluyl- α -D-ribosyl chloride and deblocking of toluylated derivatives with methanolic ammonia. Selective enzymatic phosphorylation of 12 led to its 5'-monophosphate, the latter being a moderate inhibitor of thymidylate synthase, while 12 showed moderate cytotoxicity *in vitro* against mouse leukemic cells L15178Y.

Introduction.

5-Chloro-2'-deoxyuridine exhibits cytotoxicity <u>vs</u> mouse leukemic cells L1210 and is an inhibitor of thymidylate synthase and thymidine kinase. This observation, and the fact that substitution of sulphur at the 2-position of uracil leads to an increase in acidity (i.e. decreases the pK_a for dissociation of the proton N-3, which may affect binding of the 2-thionucleosides to the above enzymes) prompted us to undertake its synthesis and to examine the properties of 5-chloro-2-thiouracil nucleosides, as well as the parent base, 5-chloro-2-thiouracil (1).

Chemistry.

Four methods were used for the synthesis of 1: (a) reaction of 5-chloro-1,3-dimethyluracil with thiourea (Scheme 1); (b) deamination of 5-chloro-2-thiocytosine (4) (Scheme 1); (c) chlorination of the p-nitrophenylethyl derivative (6) of 2-thiouracil (Scheme 2); (d) reaction of 2,5-dichloro-4-hydroxypyrimidine (9) with thiourea, which proved the most effective (60% yield) (Scheme 2).

 β - and α -5-Chloro-2-thio-2'-deoxyuridines (12 and 13) were obtained by Lewis acid catalysed condensation of TMS derivative of 1 with 2-deoxy-3,5-di-O-p-toluyl- α -D-ribosyl chloride in 1,2-dichloroethane in the presence of TiCl_a, to give a mixture of the β - and α -anomers of 2'-deoxy-3',5'-di-O-p-toluyl-5-chloro-2-thiouridine (10 and 11) with a ratio $\alpha/\beta = 1:3$ (Scheme 3). These were separated by crystallization from methanol and then purified by preparative TLC on silica gel.

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Scheme 1

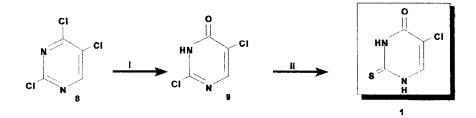
The free 5-chloro-2-thio-2'-deoxyuridine (<u>12</u>) was obtained by deblocking the p-toluyl derivative <u>10</u> with a methanolic solution of NH₃. M.p.:180-182°C; UV: λ_{max} (pH 2) 222 nm (ϵ 14300); 277 nm (ϵ 16900) λ_{max} (pH 7) 225 nm (ϵ 11800); 242 nm (ϵ 14000); 274.5 nm (ϵ 17300); λ_{max} (pH 12) 242.5 nm (ϵ 20000); 270.5 nm (ϵ 18600); ¹H NMR (500 MHz, D₂O) δ 2.33 (1 H, m, $J_{1'2}$ =6.39 Hz, 2'-H), 2.64 (1 H, m, $J_{1'2}$ =5.17 Hz, 2"-H), 3.83 (1 H, dd, $J_{4',5'}$ =4.45 Hz, 5"-H), 3.93 (1 H, dd, $J_{4',5'}$ =3.26 Hz, 5'-H), 4.11 (1 H, m, $J_{4',3'}$ =4.07 Hz, 4'-H), 4.45 (1 H, m, $J_{2',3}$ =4.54 Hz, $J_{2'',3}$ =6.46 Hz, 3'-H), 6.89 (1 H, br t, 1'-H), 8.39 (1 H, d, 6-H).

The nucleoside <u>12</u> was subjected to selective enzymatic phosphorylation with the aid of the wheat shoot nucleoside phosphotransferase system³ to give 5-chloro-2-thio-dUMP which was quantitatively converted to its parent nucleoside <u>12</u> by snake venom 5'-nucleotidase.

Antitumour Activity and Thymidylate Synthase Inhibition.

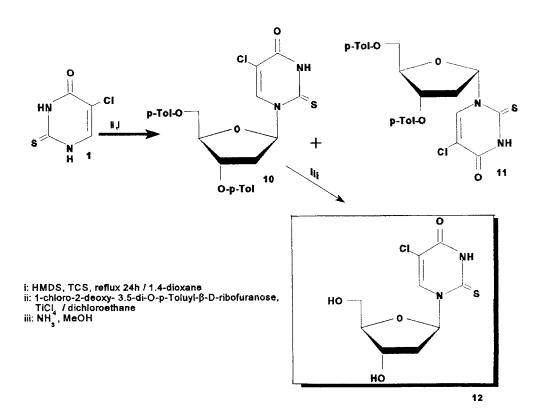
The nucleoside $\underline{12}$ was tested for antitumour activity *in vitro* with mouse leukemic cells L15178Y as previously described,⁴ and showed weaker cytotoxicity (CD₅₀ 8 x 10^{-5} M) than 5-chloro-

i: NPFE-Br, NaOH, TDA-I, 1.4-dioxane / water II: N-CI, CCI₄ , reflux, 1h iii: 0.5 N DBU in acetonitrile



i: NaOH, 45°C, 1.5h / water ii: thiourea, reflux 2h / EtOH

Scheme 2



Scheme 3

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2'-deoxyuridine (CD_{50} 5 x 10^{-7} M). To explain the mechanism of antitumour activity of $\underline{12}$, the interaction of its 5'-monophosphate with thymidylate synthase was investigated. Preparations of purified Ehrlich ascites carcinoma (EAC) and L1210 thymidylate synthase (TS) were as previously reported. Commercial L. casei TS was purified by affinity chromatography to electrophoretical homogeneity. Enzyme assays and identification of the type of inhibition were as previously described. Spectrophotometric monitoring at 340 nm of the reaction mixture containing 0.30 mM 2-thio-5-chloro-dUMP in place of dUMP, exhibited no substrate activity with L. casei thymidylate synthase. To test whether thymidylate synthase may dehalogenate this nucleoside, absorbance at 280 nm of a mixture 0.30 mM halogenated compound and 5 mM β -mercaptoethanol with TS was recorded. 2-thio-5-chloro-dUMP was not dehalogenated by L. casei enzyme. Garret et al. Peported that TS easily dehalogenates BrdUMP and IdUMP but not CldUMP.

Inhibition of Ehrlich ascites carcinoma and L1210 thymidylate synthase by 2-thio-5-chloro-dUMP was tested by varying the dUMP concentration with different concentrations of inhibitor added simultaneously to the reaction mixture. Competitive inhibition, reflected by the intersection at the ordinates of Lineweaver-Burk plots led to apparent K, values of 6.0 and 23.2 μ M for EAC and L1210 TS, respectively. The K_i of L1210 TS for 2-thio-5-chloro-dUMP was over 300-fold higher than that for 2-thio-5-fluoro-dUMP. 2-Thio-5-chloro-dUMP did not cause time-dependent inactivation of thymidylate synthase, while 2-thio-5-fluoro-dUMP is a time-dependent inactivator of TS from many sources.⁸

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